



## Using flavonoids as a therapeutic intervention against rheumatoid arthritis: The known and unknown



Zhimin Miao<sup>a,b,c,1</sup>, Yuxin Zhao<sup>a,b,c,1</sup>, Meiwan Chen<sup>a,b</sup>, Chengwei He<sup>a,b,c,\*</sup>

<sup>a</sup> State Key Laboratory of Quality Research in Chinese Medicine, Institute of Chinese Medical Sciences, University of Macau, Taipa, Macao SAR, 999078, China

<sup>b</sup> Department of Pharmaceutical Science, Faculty of Health Sciences, University of Macau, Taipa, Macao SAR 999078, China

<sup>c</sup> Guangdong-Hong Kong-Macao Joint Lab on Chinese Medicine and Immune Disease Research, University of Macau, Taipa, Macao SAR 999078, China

### ARTICLE INFO

#### Keywords:

Rheumatoid arthritis  
flavonoids  
immunoregulation  
gut-joint axis  
inflammatory responses

### ABSTRACT

Rheumatoid arthritis (RA) is an intricate autoimmune disease involved in numerous factors. Aberrant immune responses at joint sites are considered primary in the pathogenesis of RA. The complex interactions may occur between distinct immune cells, aiming at amplifying and accelerating inflammatory responses in inflamed joints. At present, gut-joint axis hypothesis holds the idea that RA originates in the gut as a result of coactions between the intestinal immune cells and dysbiotic microbiota. Dysbiosis causes intestinal inflammation and alterations in intestinal permeability, which provides a pathological basis for the transfer of activated intestinal immune cells and their products to the joints through systemic circulation or other ways. Some therapeutic options widely utilized for the treatment of RA are associated with gut-joint axis, suggesting modulation of gut-joint axis may be a promising strategy in preventing and treating RA. Flavonoids are a type of polyphenol widely existed in herbs and foods showing anti-RA potentials. However, the mechanisms by which flavonoids mitigate RA have not been well organized. In this review, we outline and discuss current understanding of the underlying mechanisms of anti-RA flavonoids through immunoregulation, gut-joint axis, and inflammatory responses, providing a reference for developing novel strategies for the treatment and prevention of RA.

### 1. Introduction

Rheumatoid arthritis (RA) refers to a chronic autoimmune disorder of unknown etiology. The capital features of RA are persistent synovitis accompanied by extra-articular organ involvement and autoantibodies including rheumatoid factor (RF) and anti-citrullinated peptide protein antibodies (ACPAs) production. Genetic background and environmental exposure are the risk factors that contribute to the development of RA [1]. Epidemiological data have shown that RA affects 0.5-1.0% of adults, and women and elderly people are susceptible to the disorder [2]. Clinically, the primary therapeutic agents of RA, including disease-modifying anti-rheumatic drugs (DMARDs), non-steroidal anti-inflammatory drugs (NSAIDs), and glucocorticoid, mitigate synovitis and systemic inflammation and relieve the pain of RA patients. However, these drugs can't produce adequate effects on the improvement of the disorder and their serious side effects including hepatorenal toxicity also restrict their clinical application [3]. Lately, biological agents are able to apply as arthritis is out-of-control or toxic effects arise with DMARDs. Owing to nasty infections and high costs, their use remains limited [3]. Fundamentally,

the application of anti-RA drugs is restricted as a consequence of the pathogenesis of RA that is not well understood.

Perturbations in immune homeostasis in joints is considered dominate in RA. It has been well reported that a variety of immune cells contribute significantly to maintaining the immune function homeostasis of host and to the pathogenesis of RA [4]. In individuals with established RA, substantial data have reported that infiltration and aberrant activation/inhibition of immune cells, such as T cells, B cells, and macrophages, are abundant in synovial tissues, which contributes to initiating and perpetuating joints inflammatory milieu [5]. The complex interplays that may occur among different immune cells, intending to expand and facilitate the inflammatory response involving joints [6]. At present, extraordinary progress has been made in the development of drugs targeting immune cells. Data came from experimental arthritis models and clinical diagnosis in individuals with RA prove that these target-specific drugs can significantly reduce pro-inflammatory cytokines, RF, ACPAs, and C-reactive protein in peripheral blood and synovial fluid [7,8]. Nevertheless, the relevance between their pharmacological effects and the reduced biomarkers of RA warrants further investigation.

The gut is generally considered to be the largest immune organ of the host owing to the maximum innate and adaptive immune cells inhab-

\* Corresponding author.

E-mail address: [chengweihe@um.edu.mo](mailto:chengweihe@um.edu.mo) (C. He).

<sup>1</sup> These authors contributed equally to this work

itation [9]. The intestinal microbiota that resides within the host gut mucous membranes comprises over 100 trillion bacteria. Compelling evidence supports that the normal gut microbe is essential for maintaining enteral and parenteral immune responses [10–12]. However, dysbiosis occurs frequently as gut microbiota is susceptible to environmental and host-related factors, thereby resulting in gut inflammation and leakiness [13], triggering bacteria penetration into the extra-intestine. Subsequently, with dysfunctional microbiota stimulation, the intestine-derived immune cells are activated and secrete pro-inflammatory cytokines, which can alter local and follow-up systemic immune responses, eventually boosting the pathogenesis of RA [14]. Mounting data conducted in different mouse models of RA and individuals with RA prove that the intestinal dysbiosis, together with intestinal inflammation and leakiness can potentially bring about arthritis [15–20]. The gut-joint axis hypothesis which holds the idea that RA begins in the intestine has recently been put forward [21]. Data reinforcing the hypothesis are that a number of intestinal innate and adaptive immune cells are amplified and activated in synovium and systemic circulation of RA patients [22]. However, few therapeutic methods have focused on gut-joint axis so far.

Flavonoids refer to a series of compounds formed by the interconnection of two phenyl rings with phenolic hydroxyl groups through the central three carbon atoms [23]. They can mainly be classified into flavone, flavanol, flavanone, flavanonol, anthocyanidin, and chalcone [24] (Fig. 1). Flavonoids with wide distribution and relatively low toxicity can be consumed safely in diet and show potent anti-inflammatory and anti-oxidant effects. Moreover, the activity of flavonoids antioxidants is in strong correlation with their chemical structures [25,26]. Flavonoids intervention with low cost is widely used in clinical treatment of various diseases, such as neurodegenerative diseases, cancers, and eye diseases [27–30]. Emerging evidence has revealed that flavonoids elicit the anti-RA potentials. Flavonoids improve symptoms of RA via multiple targets involved in immunoregulation, modulation of gut-joint axis, and inhibition of inflammatory responses.

In this review, we seek to outline the current understanding of the underlying mechanisms of anti-RA flavonoids via immune responses and gut-joint axis, providing a reference for developing novel strategies for treating and preventing RA. The literatures selected is based on a search of Pubmed between 2011 and 2021 and all doses of flavonoids presented in literatures are in accordance with the range given in the consensus document [31].

## 2. The roles of flavonoids in immune responses

It is well known that the main symptoms of RA are chronic inflammation in the joints, which leads to joint destruction [32]. Defining critical cellular subtypes and their activation/inhibition states in the inflamed joints is a key step in searching therapeutic targets for RA [4]. T cells, B cells, and macrophages have been established relevance to RA pathogenesis and are found in abundance in inflamed synovial membranes of RA patients [33–36] (Fig. 2). The mechanisms of flavonoids targeting these immune cells to ease the symptoms of RA are summarized in Table 1.

### 2.1. The roles of flavonoids in T lymphocytes

CD4<sup>+</sup> helper T (Th) cells engages in the development of RA and mainly differentiate into three subtypes with distinct immunological function. Th1 cells, originated from naïve Th cells that are mainly driven and induced by interleukin (IL)-12 produces interferon (IFN)- $\gamma$  and tumor necrosis factor (TNF)- $\alpha$  [37] and plays an important role of pro-inflammatory. IL-4 produced by basophils, eosinophils, and mature Th2 cells is the primary signal for Th2 cells lineage differentiation. Th2 cells produce anti-inflammatory cytokines like IL-4, IL-5, IL-10, and IL-13 [38]. These cytokines can induce proliferation of Th2 cells while suppressing that of Th1 cells as we as maintaining the balance of Th1/Th2. Differentiation of Th17 cells is promoted by IL-6, IL-23, and IL-1 $\beta$ . Th17

cells are mainly secreting IL-17, which contributes to cartilage and bone destruction [39]. Regulator T (Treg) cells also contribute importantly to adaptive immune responses. Induced Treg cells are the result of peripheral T cell activation in the presence of transforming growth factor (TGF)- $\beta$  costimulation and lack of pro-inflammatory cytokines [33,40]. TGF- $\beta$  and IL-10 produced by Treg cells have potent anti-inflammatory effects through blocking of T cells division and differentiation into Th17 [41]. In RA patients, the drifts of Th1/Th2 and Th17/Treg are widely existed [42,43]. Re-maintaining the homostasis of them is the crucial idea to develop anti-RA drugs.

Flavonoids derived from herbs and foods can effectively improve RA by modulating T cells in collagen-induced arthritis (CIA) and adjuvant-induced arthritis (AIA) experimental models. Acacetin is a natural flavonoid extracted from *Saussurea involucre* (Kar. et Kir.) Sch. -Bip and can significantly repress the incidence of CIA and preventing the pathological alteration by expanding Treg cells and narrowing Th1 and Th17 cells in spleen and inguinal lymph nodes [44]. Cinnamannin D1, a flavonoid derived from the food spice *Cinnamomum tamala*, elicits potent anti-RA property by regulating Th17/Treg balance in mice with CIA [45]. Quercetin, a flavonoid widely existed in fruits and vegetables, shows robust anti-arthritis effect in pre-clinical and clinical studies with unknown underlying mechanisms. Yang et al. report that quercetin can substantially yield an obvious mitigation of arthritic manifestations by decreasing the percentage of Th17 cells and increasing that of Treg cells [46]. Naringin, a well-known flavanone glycoside found in citrus fruits, can dramatically regulate Th1/Th2 balance to improve autoimmune arthritis in AIA mice [47]. Taken together, flavonoids can effectively ameliorate the symptoms of RA by regulating the homostasis of T cells.

### 2.2. The roles of flavonoids in B lymphocytes

The production of RF and ACPAs is one of the characteristics of RA, and can even occur earlier than clinical symptoms. Stimulated by antigen in synovial tissues, B lymphocytes proliferate and differentiate into a large number of plasma cells, which can synthesize and secrete antibodies as well as pro-inflammatory mediators and circulate in the peripheral blood [48,49]. Recently, B cells depletion therapy in RA has led to some surprises, however, the potential side effects can't be neglected [50–52].

Epigallocatechin gallate, a bioactive flavonoid in green tea, can effectively inhibit the migration of B cells to extravascular space, including joints and may be a promising drug for RA treatment [53]. Epigallocatechin-3-gallate is also a bioactive flavonoid derived from green tea and have a therapeutic effect in CIA rats by depleting B cells [54]. In brief, using flavonoids intervention targeting B cells is an effective approach to ameliorate the symptoms of RA.

### 2.3. The roles of flavonoids in macrophages

Macrophages that are differentiated from peripheral blood monocytes get involved in the initiation and perpetuation of inflammation, adhesion and migration of leukocyte, and degradation of matrix. It has been revealed that the inflamed synovial membrane and the cartilage-pannus junction are populated by macrophages [36], suggesting these cells may intimately associated with the pathogenetic cascade of RA. Interestingly, Macrophages are highly plastic and can switch from one phenotype to another. Stimulated by lipopolysaccharide (LPS) or IFN- $\gamma$ , macrophages can differentiate into M1 subtypes, which can produce reactive oxygen species (ROS), nitric oxide (NO), and a large number of pro-inflammatory cytokines, showing pro-inflammatory effects and subsequently causing articular cartilage injury. In the presence of IL-4 or IL-13 stimulation, macrophages can differentiate into M2 subtypes. M2 secrete anti-inflammatory cytokines and promote synovial tissues repair [36,55]. Although treatment of RA with anti-macrophage approaches is

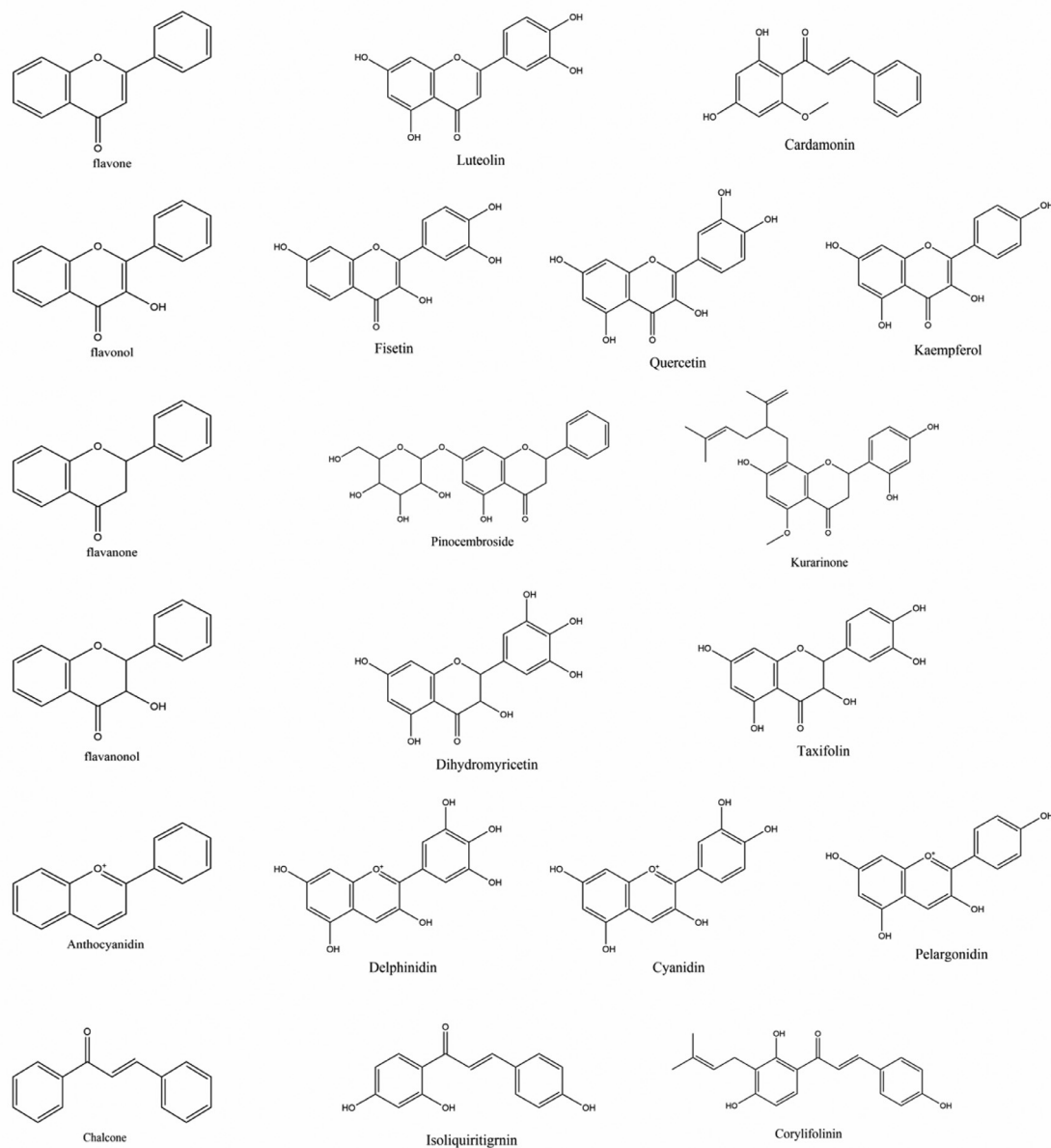


Fig. 1. The structures of flavonoids and their representative compounds.

theoretically therapeutic, regulation of macrophages polarization seems to be more promising for the treatment of RA.

Flavonoids also possess anti-RA potentials by regulating macrophages polarization. Hesperidin, a natural flavonoid widely presented in citrus fruits, can significantly relieve the symptoms of RA by inhibiting the polarization of macrophages to M1 [56]. Similarly, malvidin-3-O- $\beta$  glucoside derived from fruits can also ease RA symptoms by inhibiting the polarization of macrophages to M1 [57]. Silibinin, a natural polyphenolic flavonoid extracted from the herb *Silybum marianum*, can induce the macrophages M2 polarization to mitigate the symptoms of RA [58]. Thus, flavonoids can effectively ameliorate the symptoms of RA by modulating macrophages polarization.

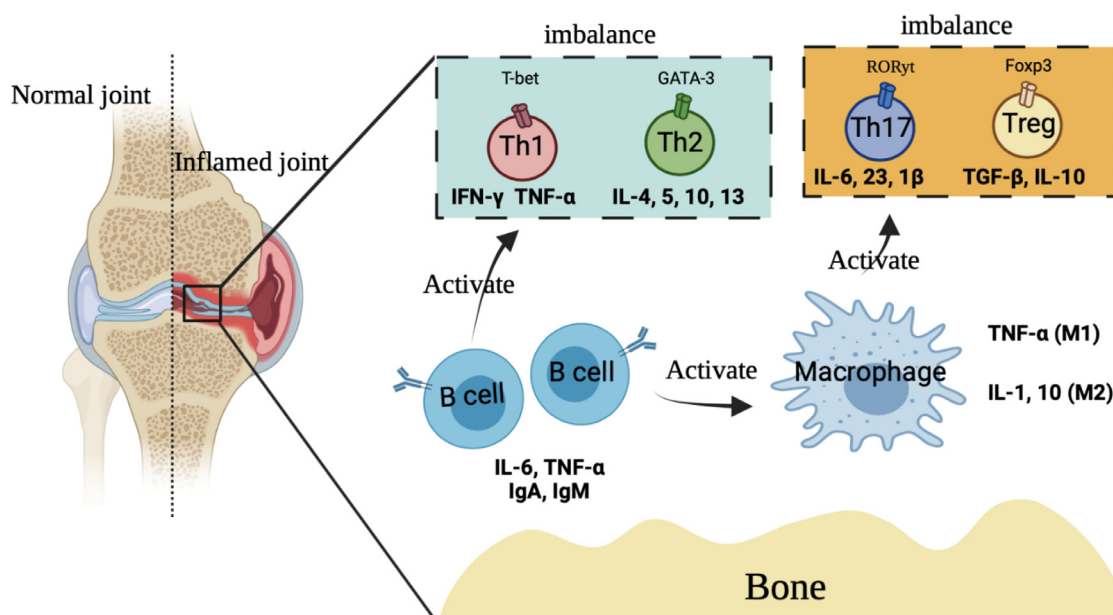
### 3. The roles of flavonoids in gut-joint axis

The gut-joint axis consists of two consecutive steps. First, the occurrence of dysbiosis accompanied by intestinal inflammation and barrier dysfunction provides the opportunity for the interactions between intestinal immune cells and dysbiotic microbiota. Second, the activated

intestinal immune cells with the abilities of pro-inflammatory cytokines production enter the joints through systemic circulation or other ways, thus leading to the pathogenesis of RA (Fig. 3). Therefore, there are two therapeutic targets for RA through gut-joint axis: (1) improve intestinal dysbiosis; (2) inhibit intestinal immune cells proliferation or trafficking.

#### 3.1. The roles of flavonoids in intestinal dysbiosis

Intestinal dysbiosis is an aberrant microbial ecological state characterized by pathobionts bloom, commensals reduction, and diversity loss. Enteric inflammation, diet, and xenobiotics are the main factors that contributes to dysbiosis [65]. Substantial data have demonstrated that dysbiosis occurs in different experimental mouse models of RA and individuals with RA. For example, Rogier et al. have demonstrated that gut microbiota undergoes significant changes in the preclinical phase of CIA, as characterized by reduced abundance of *Bacteroidetes* and raised that of *Firmicutes* [66]. Some cohort studies have also proved that great alterations in the intestinal microbial community are presented in individuals with new-onset and established RA [67–70]. Data from fecal



**Fig. 2.** The interactions among immune cells in inflamed joints. T cells, B cells, and macrophages contribute essentially to rheumatoid arthritis (RA). T cells differentiate into several subtypes that produce different cytokines, primarily including Th 1, Th2, Th17 and Treg cells. B cells with pro-inflammatory cytokines secretion and autoantibodies production can activate Th1 and macrophages. Macrophages encompass two subtypes, M1 and M2, and can activate Th17 cells.

**Table 1**

The roles of flavonoids in T cells, B cells, and macrophages.

Flavonoid name	Experimental models	Biological findings	Reference
Acacetin	Male DBA/1J mice with CIA	Treg cells ↑, Th17 cells ↓, Th1 cells ↓, regulate Th17/Treg	[44]
Cinnamtannin D1	Male DBA/1J mice with CIA; Naïve CD4 cells with TGF-β, IL-6, anti-IL-2/4, anti-IFN-γ stimulation or TGF-β, IL-2, anti-IL-4, anti-IFN-γ stimulation	Treg cells ↑, Th17 cells ↓, regulate Th17/Treg; Treg cells differentiation ↑, Th17 cells differentiation ↓	[45]
Quercetin	Peripheral blood mononuclear cells with CD3, anti-CD28, IL-23, IL-6, IL-1β, anti-IL-4, anti-IFN-γ stimulation; Female Wista rats with CIA	Th17 cells differentiation ↓; Treg cells ↑, Th17 cells ↓, regulate Th17/Treg	[46,59]
Epigallocatechin-3-gallate	Male DBA/1J mice with CIA; IFN-γ KO mice with CIA; Male SD rats with AIA	Treg cells ↑, Th17 cells ↓, regulate Th17/Treg; B cells ↓	[53,60]
Icariin	Male C57BL/6 mice with CIA	Th17 cells ↓	[61]
Anthocyanin	Male DBA/1J mice with CIA; Human CD4 <sup>+</sup> T cells with anti-CD3, anti-CD28, anti-IFN-c, anti-IL-4, IL-1β, IL-6 stimulation	Th17 cells ↓, Th17 cells differentiation ↓	[62]
Silibinin	Female Wista rats with CIA; Naïve CD4 <sup>+</sup> CD62L <sup>+</sup> T cells with anti-CD3, anti-CD28, TGF-β, IL-6, anti-IFN-γ, anti-IL-4 stimulation; RAW264.7 cells with LPS, IFN-γ, or IL-4 stimulation	Th17 cells ↓, Th17 cells differentiation ↓; Macrophage M2 polarization ↑	[58]
Oroxylin	Male DBA/1 mice with CIA	Treg cells ↑, Th17 cells ↓, regulate Th17/Treg ratio	[63]
Naringenin	Male DBA/1J mice with CIA	Th1 cells ↓, Th17 cells ↓	[64]
Naringin	Female Balb/c mice with AIA	Th1 cells ↑, Treg cells ↑, Th2 cells ↓	[47]
Kurarinone	Male DBA/1 mice with CIA	Th1 cells ↓, Th17 cells ↓	[64]
Hesperidin	RAW264.7 cells with LPS stimulation	Macrophage M1 polarization ↓	[56]
Malvidin-3-O-β glucoside	RAW264.7 cells with LPS stimulation	Macrophage M1 polarization ↓	[57]

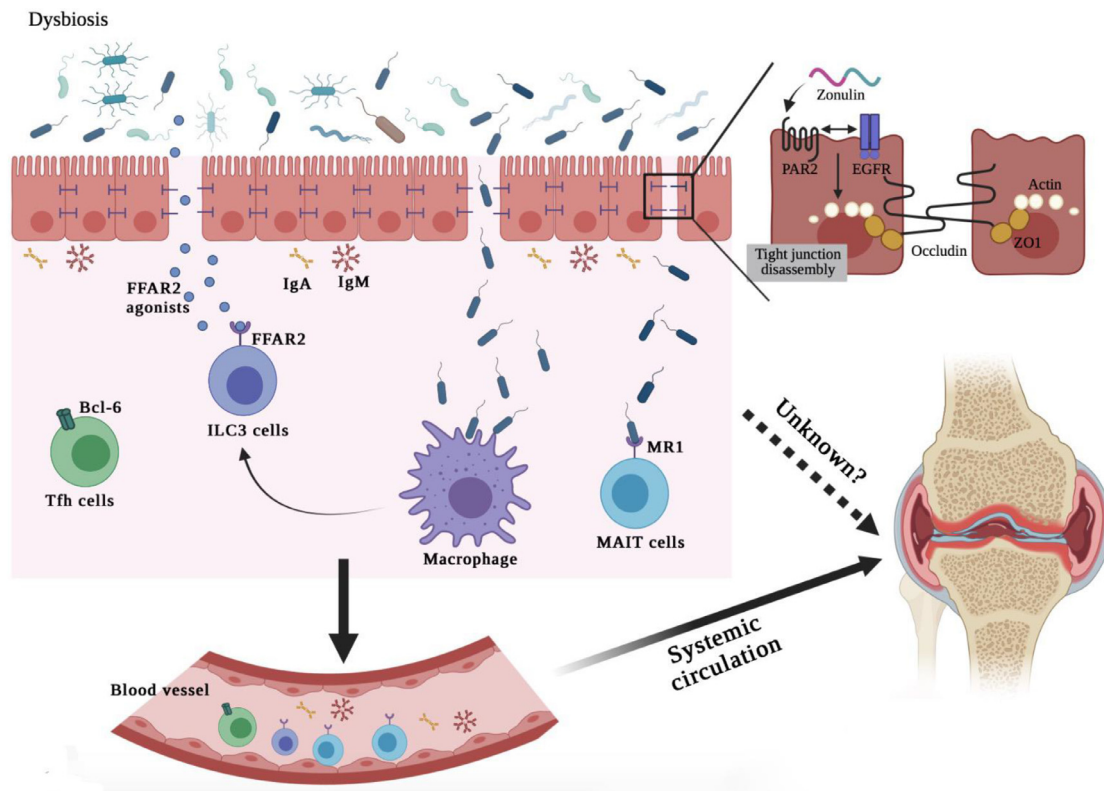
CIA, collagen-induced arthritis; AIA, adjuvant-induced arthritis

transplantation and antibiotics treatment experiments in mice prove the potential cause relationship between gut microbiota and RA. Maeda et al. have shown that colonization of SKG mice with faecal microbiota from patients with RA shows an elevated susceptibility to arthritis [71]. Jubair et al. have reported that depleting the microbiota of the mice with a cocktail of antibiotics before the induction of CIA leads to a significant reduction in the severity of disorder [72].

In light of the gut-joint axis, intestinal inflammation and gut barrier dysfunction are also major contributors in the development of RA. The occurrence of dysbiosis can trigger and perpetuate the aberrant activation and expansion of intestinal innate immune cells, therefore leading to intestinal inflammation [73]. The link between dysbiosis

and gut barrier is well conducted. The presence of normal gut microbiota can induce the differentiation of RORγt<sup>+</sup>NKp46<sup>+</sup> natural killer-like cells with IL-22 production, which can promote gut integrity and inhibit bacterial infiltration [74]. However, perturbations in gut microbiota disturb the process, causing the gut barrier dysfunction [75]. Moreover, elevated serum zonulin concentrations is associated with intestinal dysbiosis [76]. Zonulin, a biomarker of gut permeability, can initiate proteinase-activated receptor 2 (PAR2)-dependent transactivation of epidermal growth factor receptor (EGFR), therefore causing tight junction disassembly [77]. Although no direct evidence shows the correlations among intestinal inflammation, gut barrier and RA, a cohort study have reported a considerable association between RA and inflam-





**Fig. 3.** The pathogenesis of rheumatoid arthritis (RA) through gut-joint axis. Intestinal dysbiosis accompanied with intestinal inflammation and gut barrier dysfunction leads to the interactions between intestinal immune cells and aberrant microbes. Subsequently, the activated intestinal immune cells such as group 3 innate lymphoid (ILC3) cells, mucosa-associated invariant T (MAIT) cells, and T follicular helper (Tfh) cells traffic to the joints via systemic circulation, therefore promoting the pathogenesis of RA.

matory bowel disease (IBD) [78]. IBD is also an autoimmune disease with similar pathogenic mechanisms of RA and characterized by chronic intestinal inflammation and gut leakiness, suggesting that intestinal inflammation and gut barrier dysfunction have a potential contributor to the development RA. Taken together, improving intestinal dysbiosis is a possible therapeutic method to improve the development of RA.

At present, Aa et al. reported that Kaempferol, a natural flavonoid found in many edible herbs, elicit anti-arthritis activities by re-balancing dysbiosis in CIA mice [79]. Due to similar pathogenic mechanisms between RA and IBD, some studies have revealed that some flavonoids can effectively alleviate the symptoms of IBD, which can provide some hints for RA therapies using flavonoids intervention. Ren et al. have reported that acacetin can significantly improve the clinical symptoms of dextran sulfate sodium (DSS)-induced colitis in mice by modulating gut microbiota and inhibiting intestinal inflammation [80]. Zhu et al. have revealed that baicalin, a flavonoid extracted from the root of herb *Astragalus Radix*, can protect the rats against trinitrobenzene sulphonic acid (TNBS)-induced colitis by regulating gut microbiota, especially increasing the *Butyrivomonas* abundance [81]. Phloretin, a flavonoid found in pears of fruits, shows anti-IBD effects by remodeling gut microbiota, especially enriching *Lactobacillus* [82]. Pinocembrin, a plant-derived flavonoid with anti-inflammation effects, can improve the severity of DSS-induced colitis in mice by regulating gut microbiota and repairing gut barrier [83]. Taken together, data presented above suggest that flavonoids may be a type of promising drugs for RA treatment, which warrants further development [84].

### 3.2. The roles of flavonoids in intestinal immune cells proliferation and trafficking

Intestinal immune cells mainly include Group 3 innate lymphoid (ILC3) cells, mucosa-associated invariant T (MAIT) cells and intestinal

T follicular helper (Tfh) cells. ILC3 cells are primarily situated at epithelial barrier surfaces and involved in response to the extra-intestinal pathogens and in maintaining intestinal homeostasis. With commensal microbes and their products (such as free fatty acid receptor 2 agonism) stimulation, ILC3 cells are activated and expanded in the intestine and produce pro-inflammatory mediators including IL-17 and IL-22 [85,86]. In a CIA mouse model, ILC3 cells are significantly amplified in flamed joints [87]. In individuals with early RA, the number of ILC3 cells are significantly elevated in inguinal lymph node biopsy [88]. MAIT cells, mainly situated at mucosal and epithelial barrier, are innate-like T cells that bridge gut microbiota and intestinal immune responses [89]. They are activated by certain bacteria and subsequently produce pro-inflammation mediators. Blunted MAIT cells frequency in systemic circulation and elevated of that in inflamed tissues are common features in various autoimmune diseases [89]. In the context of RA, MAIT cells are abundant in the synovial fluid and high levels of pro-inflammation mediators can promote the migration of MAIT cells from blood to the joints [90]. Tfh cells are a subset of activated CD4<sup>+</sup> T cells which can assist the formation and maintenance of germinal centers (GC). Tfh promote B lymphocytes differentiation and autoantibodies production by secreting IL-21 [91]. The migration of Tfh cells from the intestine to the joints has been well conducted [92]. A cohort study has demonstrated that a higher percentage of circulating Tfh cells and higher serum level of IL-21 in RA patients [93]. At present, it is well understood that these cells can enter the joints through systemic circulation. Nevertheless, there may have other ways promote the communication between gut and joints, which needs further investigation. Taken together, inhibiting the proliferation and migration of intestinal immune cells is also a therapeutic way for the treatment of RA.

Although no research targeting the intestinal immune cells to improve RA using flavonoids intervention has been reported, some investigators have demonstrated that flavonoids can have a therapeutic role

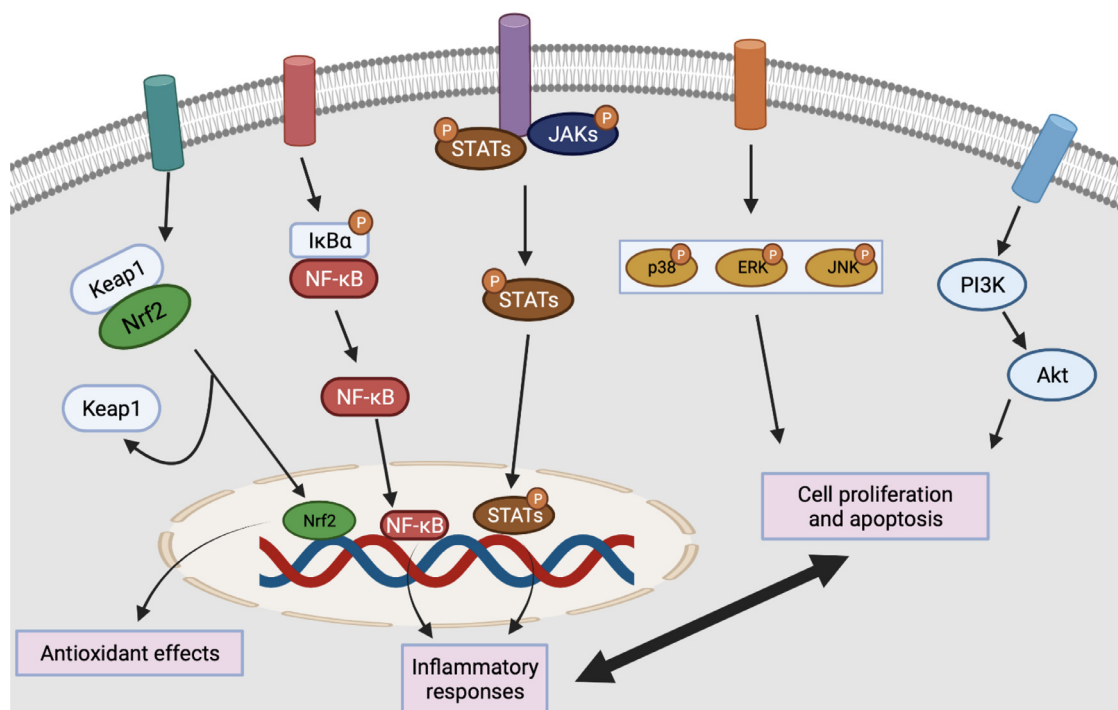


Fig. 4. Therapeutic signaling pathways of flavonoids

in some autoimmune diseases through intestinal immune cells, which is also beneficial for the anti-RA flavonoid drugs development. Yang et al. have revealed that baicalin attenuates lupus autoimmunity by inhibiting Tfh cells differentiation and pro-inflammation cytokines production [94].

#### 4. The roles of flavonoids in inflammatory mediators

Inflammatory mediators including histamine, cytokines, chemokines, integrins, and proteases that are generated by immune cells secretion and genetic expression are responsible for bone erosion and inflammation at synovial sites and for the progress of RA. Nowadays, balancing and resolving inflammation are regarded as promising strategies for RA. Flavonoids are well-known for their robust anti-inflammatory effects and overwhelming evidence shows that flavonoids lighten the symptoms of RA through inhibiting local and systemic inflammation responses. Consumption of cinnamtannin D1 can significantly increase TGF- $\beta$  and IL-10 levels while decrease those of IL-6, IL-17 and IL-1 $\beta$  in serum of CIA mice [45]. Hesperidin treatment can effectively suppress the expression of matrix metalloproteinases (MMP) in LPS-induced FLS cells [54]. Malvidin-3-O- $\beta$  glucoside treatment can substantially inhibit the secretion of IL-1, IL-6, and NO in LPS-induced macrophages [57]. Icaritin addition can dramatically inhibit the expression of  $\beta$ 3 integrin and MMP9 in bone marrow-derived macrophage cells [61]. Administration of oroxylin A can memorably decrease the serum levels of IL-1 $\beta$ , IL-6, IL-17, and TNF- $\alpha$  [63]. Kurarinone can also markedly decrease the serum and paw tissues levels of IFN- $\gamma$ , TNF- $\alpha$ , IL-6, and IL-17A. Taken together, flavonoids targeting inflammatory mediators can effectively improve the development of RA.

#### 5. Therapeutic signaling pathways of flavonoids

The development of RA is an intricate process involving many signaling pathways, primarily including PI3K/Akt, MAPK, NF- $\kappa$ B, STAT, and Nrf2 signaling pathways (Fig. 4).

##### 5.1. The roles of flavonoids in PI3K/Akt signaling pathway

Synovial hyperplasia and inflammation are considered to be the pathological features of RA. In the context of RA, fibroblast-like synovial (FLS) cells can secrete a plenty of inflammatory cytokines and continuously stimulate FLS cells, resulting in uncontrolled proliferation. PI3K/Akt signaling pathway is deemed to be a bridge between proliferation and apoptosis of FLS cells. In RA-FLS cells, PI3K and Akt are highly expressed, and have an effect on the excessive migration of FLS cells. Moreover, lots of inflammatory cytokines such as IL-17 and IL-21 can promote the inflammatory proliferation of FLS cells by inducing and triggering PI3K [95]. Previous studies have demonstrated that LY294002, an inhibitor of PI3K, can dramatically improve synovial hyperplasia and inflammation in mice with CIA and AIA [56], suggesting targeting PI3K/Akt signaling pathway is an effective mean to inhibit the progress of RA. Nowadays, increasing literatures have reported that flavonoids can significantly improve the development of RA in experimental mouse models through PI3K/Akt signaling pathway. Qi et al. have reported that hesperidin can significantly inhibit the expression of PI3K and p-Akt in M1 macrophages and FLS cells. Moreover, upon LY294002 intervention, the anti-arthritis effects of hesperidin are dampened in AIA mice [56]. Liu et al. have revealed that epigallo-catechin-3-gallate can suppress the expression of p-Akt in B cells of CIA rats, therefore repressing the development of RA [54].

##### 5.2. The roles of flavonoids in MAPK signaling pathway

MAPK signaling pathway including extracellular signal-related kinase (ERK-1/2), p38 (p38 $\alpha$ / $\beta$ / $\gamma$ / $\delta$ ), c-Jun N-terminal kinase (JNK-1/2/3), and ERK5 that regulates a series of cytokines, chemokines, and enzymes. MAPK signaling pathway is hyperactive in synovial tissues and leads to persistent inflammation and abnormal hyperplasia [96,97]. The selective inhibitors targeting MAPK signaling pathway have been proposed to be an effective approach for the treatment of RA [98]. Similarly, the MAPK signaling pathway is inhibited accompanied with reduced symptoms of RA upon flavonoids treatment. Sun et al. have demonstrated that the flavonoids, extracted from the herb *Flemingia philippinen-*

sis, can improve the symptoms of RA by inhibiting the expression of p-ERK1, p-p38, and p-JNK in paw tissues of CIA mice [99]. Zhai et al. have reported that liquiritin, a flavonoid extracted from the herb *Glycyrrhiza uralensis*, can significantly inhibit the IL-1 $\beta$ -induced RA-FLS proliferation by down-regulating p-p38, and p-JNK [100].

### 5.3. The roles of flavonoids in NF- $\kappa$ B and STAT signaling pathways

NF- $\kappa$ B and STAT also contribute significantly to RA. When NF- $\kappa$ B and STAT are activated, they then shift to the nucleus and gets involved in the regulation of inflammatory response, cell proliferation and apoptosis [101,102]. NF- $\kappa$ B and STAT are profoundly and sustainably activated in CIA and AIA mice, which are improved by flavonoids. Bai et al. have reported that baicalin can alleviate CIA in rats and repress RA-FLS proliferation by down-regulating nuclear p65 expression [103]. Bao et al. have reported that Genkwanin, a flavonoid isolated from the herb *Daphne genkwa*, exerts anti-RA effects by down-regulating the expression of p-STAT3 and p-NF- $\kappa$ B in paw tissues of AIA mice [101].

### 5.4. The roles of flavonoids in NRF2 signaling pathways

Oxidative stress is involved in the pathogenesis and pathological process of RA. Upon activating Nrf2 signaling pathway, various antioxidant enzymes (such as heme oxygenase-1, HO-1) are released to regulate the oxidative stress state of RA [104]. It has been well conducted that Nrf2-deficiency increases susceptibility to RA [105] and the mice with Nrf2-knockout background shows higher levels of pro-inflammatory mediators than their wild-type littermates [106]. The Nrf2 signaling pathway is activated accompanied with reduced symptoms of RA upon flavonoids treatment. Karatas et al. have reported that epigallocatechin 3-gallate had anti-arthritis effects by up-regulating the expression of Nrf2 and HO1 in joint tissues of CIA rats [107]. Su et al. have reported that calycosin, a flavonoid isolated from the herb *Astragali Radix*, can suppress the expression of pro-inflammatory mediators in RA-FLS by up-regulating the expression of Nrf2 and HO1 [108].

## 6. Concluding remarks

Nowadays, gut microbiota has been a hot target in a variety of diseases. Alterations in the composition of gut microbiota have been observed in mice with CIA and individuals with established RA [21], suggesting gut microbiota may also get involved in the occurrence and progression of RA. With the advent of metagenome sequencing and the development of germ-free and humanized mouse models, the association between gut microbiota and RA has been well understood. In light of these existing data, the concepts of gut-joint axis are described, replenishing the pathogenesis of RA.

The main drawback of flavonoids is that they take effect slowly and need to be taken for a long time, which is also related to their poor absorption. Most of flavonoids are taken orally and can encounter commensal bacteria in the small and large intestine. These microbes collectively encode 150-fold-more genes than human genome [109], showing a rich enzyme repository with drug-metabolizing potentials. In fact, there are quite a number of flavonoids metabolized by gut microbiota, and the metabolites possess better absorption and more active pharmacological activities than their parent drugs. Take scutellarin, a flavonoid with high effectiveness in clinic, for example. The oral bioavailability of scutellarin, is exceptionally low. In healthy volunteers and rats, the oral bioavailability of scutellarin was found to be merely 2.2% and 0.67% respectively [110]. The physiological effects of scutellarin are in notable contrast to its poor bioavailability. Although scutellarin seems to be absorbed in the form of scutellarein, no aglycone but isoscutellarin is detected in the portal vein plasma and the plasma concentrations of isoscutellarin exceed that of scutellarin by about 30-fold. Furthermore, gut microbiota also gets involved in the process of isoscutellarin

transformation and isoscutellarin shows more excellent pharmacological effects than scutellarin [110]. Taken together, focus on flavonoids gut microbiota-derived metabolites seems to be a promising method to search for novel drugs.

Although using flavonoids as a therapeutic intervention against RA is at a very initial stage and still needs a lot of pre-clinical and clinical data, researches are shedding light on flavonoids in potential clinical treatment and prevention of RA. Further studies are required specifically to define the exact step of gut-joint axis (especially the intestinal mucosal-derived immune cells trafficking step) during the pathogenesis of RA and to develop flavonoids for RA treatment with the focus on the interplays with flavonoids and gut microbiota.

## Declaration of competing interest

No conflicts of interest in this article

## Acknowledgements

This work was supported by the Macao Science and Technology Development Fund, Macao SAR (0024/2020/A1), the Research Fund of the University of Macau (MYRG2018-00176-ICMS) and the 2020 Guangdong Provincial Science and Technology Innovation Strategy Special Fund (Guangdong-Hong Kong-Macau Joint Lab) (2020B1212030006).

## References

- [1] IB McInnes, G. Schett, The pathogenesis of rheumatoid arthritis, *N Engl J Med* 365 (23) (2011) 2205–2219, doi:10.1056/NEJMra1004965.
- [2] DL Scott, F Wolfe, TW. Huizinga, Rheumatoid arthritis, *Lancet* 376 (9746) (2010) 1094–1108, doi:10.1016/S0140-6736(10)60826-4.
- [3] HJ Li, CT Zhang, H Du, et al., Chemical Composition of Bawei Longzuan Granule and Its Anti-Arthritic Activity on Collagen-Induced Arthritis in Rats by Inhibiting Inflammatory Responses, *Chem Biodivers* 16 (9) (2019) e1900294, doi:10.1002/cbdv.201900294.
- [4] F Zhang, K Wei, K Slowikowski, et al., Defining inflammatory cell states in rheumatoid arthritis joint synovial tissues by integrating single-cell transcriptomics and mass cytometry, *Nat Immunol* 20 (7) (2019) 928–942, doi:10.1038/s41590-019-0378-1.
- [5] Y He, Y Huang, C Mai, et al., The immunomodulatory role of PDEs inhibitors in immune cells: therapeutic implication in rheumatoid arthritis, *Pharmacol Res* 161 (2020) 105134, doi:10.1016/j.phrs.2020.105134.
- [6] CM Weyand, JJ. Goronzy, The immunology of rheumatoid arthritis, *Nat Immunol* 22 (1) (2021) 10–18, doi:10.1038/s41590-020-00816-x.
- [7] ST Law, PC. Taylor, Role of biological agents in treatment of rheumatoid arthritis, *Pharmacol Res* 150 (2019) 104497, doi:10.1016/j.phrs.2019.104497.
- [8] A Kubota, T Suguro, A Nakajima, et al., Effect of biological agents on synovial tissues from patients with rheumatoid arthritis, *Mod Rheumatol* 30 (2) (2020) 282–286, doi:10.1080/14397595.2019.1583783.
- [9] H Xu, H Zhao, D Fan, et al., Interactions between Gut Microbiota and Immunomodulatory Cells in Rheumatoid Arthritis, *Mediators Inflamm* 2020 (2020) 1430605, doi:10.1155/2020/1430605.
- [10] AI Catrina, KD Deane, JU. Scher, Gene, environment, microbiome and mucosal immune tolerance in rheumatoid arthritis, *Rheumatology* 55 (3) (2016) 391–402, doi:10.1093/rheumatology/keu469.
- [11] JU Scher, V Joshua, A Artacho, et al., The lung microbiota in early rheumatoid arthritis and autoimmunity, *Microbiome* 4 (1) (2016) 60, doi:10.1186/s40168-016-0206-x.
- [12] Y Maeda, K. Takeda, Host-microbiota interactions in rheumatoid arthritis, *Exp Mol Med* 51 (12) (2019) 1–6, doi:10.1038/s12276-019-0283-6.
- [13] J Henao-Mejia, E Elinav, C Jin, et al., Inflammasome-mediated dysbiosis regulates progression of NAFLD and obesity, *Nature* 482 (7384) (2012) 179–185, doi:10.1038/nature10809.
- [14] M Levy, AA Kolodziejczyk, CA Thaiss, Elinav E. Dysbiosis and the immune system, *Nat Rev Immunol* 17 (4) (2017) 219–232, doi:10.1038/nri.2017.7.
- [15] J Chen, K Wright, JM Davis, et al., An expansion of rare lineage intestinal microbes characterizes rheumatoid arthritis, *Genome Med* 8 (1) (2016) 43, doi:10.1186/s13073-016-0299-7.
- [16] Y Maeda, T Kurakawa, E Umemoto, et al., Dysbiosis Contributes to Arthritis Development via Activation of Autoreactive T Cells in the Intestine, *Arthritis Rheumatol* 68 (11) (2016) 2646–2661, doi:10.1002/art.39783.
- [17] J. Inamo, Non-causal association of gut microbiome on the risk of rheumatoid arthritis: a Mendelian randomisation study, *Ann Rheum Dis* 80 (7) (2021) e103, doi:10.1136/annrheumdis-2019-216565.
- [18] G Horta-Baas, MDS Romero-Figueroa, AJ Montiel-Jarquín, et al., Intestinal Dysbiosis and Rheumatoid Arthritis: A Link between Gut Microbiota and the Pathogenesis of Rheumatoid Arthritis, *J Immunol Res* 2017 (2017) 4835189, doi:10.1155/2017/4835189.



- [19] F Salem, N Kindt, JR Marchesi, et al., Gut microbiome in chronic rheumatic and inflammatory bowel diseases: Similarities and differences. *United, European Gastroenterol J* 7 (8) (2019) 1008–1032, doi:10.1177/2050640619867555.
- [20] D Alpizar-Rodriguez, TR Lesker, A Gronow, et al., Prevotella copri in individuals at risk for rheumatoid arthritis. *Ann Rheum Dis* 78 (5) (2019) 590–593, doi:10.1136/annrheumdis-2018-214514.
- [21] MM Zaiss, HJ Joyce Wu, D Mauro, G Schett, F Ciccia, The gut-joint axis in rheumatoid arthritis. *Nat Rev Rheumatol* 17 (4) (2021) 224–237, doi:10.1038/s41584-021-00585-3.
- [22] E May, E Märker-Hermann, BM Wittig, et al., Identical T-cell expansions in the colon mucosa and the synovium of a patient with enterogenic spondyloarthritis. *Gastroenterology* 119 (6) (2000) 1745–1755, doi:10.1053/gast.2000.20173.
- [23] E Corradini, P Foglia, P Giansanti, et al., Flavonoids: chemical properties and analytical methodologies of identification and quantitation in foods and plants. *Nat Prod Res* 25 (5) (2011) 469–495, doi:10.1080/14786419.2010.482054.
- [24] MA Lila, Anthocyanins and Human Health: An In Vitro Investigative Approach. *J Biomed Biotechnol* 2004 (5) (2004) 306–313, doi:10.1155/S111072430440401X.
- [25] I Glevitzky, GA Dumitrel, M Glevitzky, et al., Statistical Analysis of the Relationship Between Antioxidant Activity and the Structure of Flavonoid Compounds. *Revista de Chimie* 70 (9) (2019) 3103–3107, doi:10.37358/RC.19.9.7497.
- [26] A Pallag, SG Bungau, DM Tit, et al., Comparative study of polyphenols, flavonoids, and chlorophylls in *Equisetum Arvense* T. populations. *Rev Chim* 67 (3) (2016) 530–533.
- [27] T Behl, G Kaur, A Sehgal, et al., Flavonoids, the Family of Plant-derived Antioxidants making inroads into Novel Therapeutic Design against IR-induced Oxidative Stress in Parkinson's Disease. *Curr Neuropharmacol* (2021), doi:10.2174/1570159x19666210524152817.
- [28] R Makkar, T Behl, S Bungau, et al., Nutraceuticals in Neurological Disorders. *Int J Mol Sci* 21 (12) (2020) 4424, doi:10.3390/ijms21124424.
- [29] VS Sivasankarapillai, R Madhu Kumar Nair, A Rahdar, et al., Overview of the anticancer activity of withaferin A, an active constituent of the Indian ginseng *Withania somnifera*. *Environ Sci Pollut Res Int* 27 (21) (2020) 26025–26035, doi:10.1007/s11356-020-09028-0.
- [30] S Bungau, MM Abdel-Daim, DM Tit, et al., Health Benefits of Polyphenols and Carotenoids in Age-Related Eye Diseases. *Oxid Med Cell Longev* 2019 (2019) 9783429, doi:10.1155/2019/9783429.
- [31] M Heinrich, G Appendino, T Efferth, et al., Best practice in research - Overcoming common challenges in phytopharmacological research. *J Ethnopharmacol* 246 (2020) 112230, doi:10.1016/j.jep.2019.112230.
- [32] F Wolfe, DM Mitchell, JT Sibley, et al., The mortality of rheumatoid arthritis. *Arthritis Rheum* 37 (4) (1994) 481–494, doi:10.1002/art.1780370408.
- [33] AM Gizinski, DA Fox, T cell subsets and their role in the pathogenesis of rheumatic disease. *Curr Opin Rheumatol* 26 (2) (2014) 204–210, doi:10.1097/BOR.0000000000000036.
- [34] CC Reparón-Schuijt, WJ van Esch, C van Kooten, et al., Secretion of anticitrulline-containing peptide antibody by B lymphocytes in rheumatoid arthritis. *Arthritis Rheum* 44 (1) (2001) 41–47, doi:10.1002/1529-0131(200101)44:1<41::AID-ANR6>3.0.CO;2-0.
- [35] D Mulherin, O Fitzgerald, B Bresnihan, Synovial tissue macrophage populations and articular damage in rheumatoid arthritis. *Arthritis Rheum* 39 (1) (1996) 115–124, doi:10.1002/art.1780390116.
- [36] RW Kinne, R Bräuer, B Stuhlmeier, E Palombo-Kinne, GR Burmester, Macrophages in rheumatoid arthritis. *Arthritis Res* 2 (3) (2000) 189–202, doi:10.1186/ar86.
- [37] M Ren, M Kazemian, M Zheng, et al., Transcription factor p73 regulates Th1 differentiation. *Nat Commun* 11 (1) (2020) 1475, doi:10.1038/s41467-020-15172-5.
- [38] F Van Gool, MLT Nguyen, MR Mumbach, et al., A Mutation in the Transcription Factor Foxp3 Drives T Helper 2 Effector Function in Regulatory T Cells. *Immunity* 50 (2) (2019) 362–377.e6, doi:10.1016/j.immuni.2018.12.016.
- [39] K Yasuda, Y Takeuchi, K Hirota, The pathogenicity of Th17 cells in autoimmune diseases. *Semin Immunopathol* 41 (3) (2019) 283–297, doi:10.1007/s00281-019-00733-8.
- [40] ME Free, DO Bunch, JA McGregor, et al., Patients with antineutrophil cytoplasmic antibody-associated vasculitis have defective Treg cell function exacerbated by the presence of a suppression-resistant effector cell population. *Arthritis Rheum* 65 (7) (2013) 1922–1933, doi:10.1002/art.37959.
- [41] Y Wang, S Chen, K Du, et al., Traditional herbal medicine: Therapeutic potential in rheumatoid arthritis. *J Ethnopharmacol* 279 (2021) 114368, doi:10.1016/j.jep.2021.114368.
- [42] Y Zhang, Y Zhang, W Gu, L He, B. Sun, Th1/Th2 cell's function in immune system. *Adv Exp Med Biol* 841 (2014) 45–65, doi:10.1007/978-94-017-9487-9\_3.
- [43] P Fasching, M Stradner, W Graninger, et al., Therapeutic Potential of Targeting the Th17/Treg Axis in Autoimmune Disorders. *Molecules* 22 (1) (2017) 134, doi:10.3390/molecules22010134.
- [44] L Liu, J Yang, B Zu, et al., Acacetin regulated the reciprocal differentiation of Th17 cells and Treg cells and mitigated the symptoms of collagen-induced arthritis in mice. *Scand J Immunol* 88 (4) (2018) e12712, doi:10.1111/sji.12712.
- [45] C Shi, H Zhang, X Wang, et al., Cinnamtannin D1 attenuates autoimmune arthritis by regulating the balance of Th17 and treg cells through inhibition of aryl hydrocarbon receptor expression. *Pharmacol Res* 151 (2020) 104513, doi:10.1016/j.phrs.2019.104513.
- [46] Y Yang, X Zhang, M Xu, et al., Quercetin attenuates collagen-induced arthritis by restoration of Th17/Treg balance and activation of Heme Oxygenase 1-mediated anti-inflammatory effect. *Int Immunopharmacol* 54 (2018) 153–162, doi:10.1016/j.intimp.2017.11.013.
- [47] SF Ahmad, KM Zoheir, Abdel-Hamid HE, et al., Amelioration of auto-immune arthritis by naringin through modulation of T regulatory cells and Th1/Th2 cytokines. *Cell Immunol* 287 (2) (2014) 112–120, doi:10.1016/j.cellimm.2014.01.001.
- [48] M Volkov, KA van Schie, D van der Woude, Autoantibodies and B Cells: The ABC of rheumatoid arthritis pathophysiology. *Immunol Rev* 294 (1) (2020) 148–163, doi:10.1111/imr.12829.
- [49] JL Barnas, RJ Looney, JH. Anolik, B cell targeted therapies in autoimmune disease. *Curr Opin Immunol* 61 (2019) 92–99, doi:10.1016/j.coi.2019.09.004.
- [50] DSW Lee, OL Rojas, JL. Gommerman, B cell depletion therapies in autoimmune disease: advances and mechanistic insights. *Nat Rev Drug Discov* 20 (3) (2021) 179–199, doi:10.1038/s41573-020-00092-2.
- [51] KA Sacco, RS. Abraham, Consequences of B-cell-depleting therapy: hypogammaglobulinemia and impaired B-cell reconstitution. *Immunotherapy* 10 (8) (2018) 713–728, doi:10.2217/imt-2017-0178.
- [52] TA Barr, P Shen, S Brown, et al., B cell depletion therapy ameliorates autoimmune disease through ablation of IL-6-producing B cells. *J Exp Med* 209 (5) (2012) 1001–1010, doi:10.1084/jem.20111675.
- [53] K Kawai, NH Tsuno, J Kitayama, et al., Catechin inhibits adhesion and migration of peripheral blood B cells by blocking CD11b. *Immunopharmacol Immunotoxicol* 33 (2) (2011) 391–397, doi:10.3109/08923973.2010.522195.
- [54] D Liu, P Li, S Song, et al., Pro-apoptotic effect of epigallocatechin-3-gallate on B lymphocytes through regulating BAFF/PI3K/Akt/mTOR signaling in rats with collagen-induced arthritis. *Eur J Pharmacol* 690 (1-3) (2012) 214–225, doi:10.1016/j.ejphar.2012.06.026.
- [55] WP Arend, M Malyak, CJ Guthridge, et al., Interleukin-1 receptor antagonist: role in biology. *Annu Rev Immunol* 16 (1998) 27–55, doi:10.1146/annurev.immunol.16.1.27.
- [56] W Qi, C Lin, K Fan, et al., Hesperidin inhibits synovial cell inflammation and macrophage polarization through suppression of the PI3K/AKT pathway in complete Freund's adjuvant-induced arthritis in mice. *Chem Biol Interact* 306 (2019) 19–28, doi:10.1016/j.cbi.2019.04.002.
- [57] A Decendit, M Mamani-Matsuda, V Aumont, et al., Malvidin-3-O- $\beta$  glucoside, major grape anthocyanin, inhibits human macrophage-derived inflammatory mediators and decreases clinical scores in arthritic rats. *Biochem Pharmacol* 86 (10) (2013) 1461–1467, doi:10.1016/j.bcp.2013.06.010.
- [58] WW Tong, C Zhang, T Hong, et al., Silibinin alleviates inflammation and induces apoptosis in human rheumatoid arthritis fibroblast-like synoviocytes and has a therapeutic effect on arthritis in rats. *Sci Rep* 8 (1) (2018) 3241, doi:10.1038/s41598-018-21674-6.
- [59] HR Kim, BM Kim, JY Won, et al., Quercetin, a Plant Polyphenol, Has Potential for the Prevention of Bone Destruction in Rheumatoid Arthritis. *J Med Food* 22 (2) (2019) 152–161, doi:10.1089/jmf.2018.4259.
- [60] SY Lee, YO Jung, JG Ryu, et al., Epigallocatechin-3-gallate ameliorates autoimmune arthritis by reciprocal regulation of T helper-17 regulatory T cells and inhibition of osteoclastogenesis by inhibiting STAT3 signaling. *J Leukoc Biol* 100 (3) (2016) 559–568, doi:10.1189/jlb.3A0514-261RR.
- [61] L Chi, W Gao, X Shu, et al., A natural flavonoid glucoside, icariin, regulates Th17 and alleviates rheumatoid arthritis in a murine model. *Mediators Inflamm* 2014 (2014) 392062, doi:10.1155/2014/392062.
- [62] HK Min, SM Kim, SY Baek, et al., Anthocyanin Extracted from Black Soybean Seed Coats Prevents Autoimmune Arthritis by Suppressing the Development of Th17 Cells and Synthesis of Proinflammatory Cytokines by Such Cells, via Inhibition of NF- $\kappa$ B. *PLoS One* 10 (11) (2015) e0138201, doi:10.1371/journal.pone.0138201.
- [63] YL Wang, JM Gao, LZ. Xing, Therapeutic potential of Oroxylin A in rheumatoid arthritis. *Int Immunopharmacol* 40 (2016) 294–299, doi:10.1016/j.intimp.2016.09.006.
- [64] YR Li, DY Chen, CL Chu, et al., Naringenin inhibits dendritic cell maturation and has therapeutic effects in a murine model of collagen-induced arthritis. *J Nutr Biochem* 26 (12) (2015) 1467–1478, doi:10.1016/j.jnutbio.2015.07.016.
- [65] M Levy, AA Kolodziejczyk, CA Thaiss, et al., Dysbiosis and the immune system. *Nat Rev Immunol* 17 (4) (2017) 219–232, doi:10.1038/nri.2017.7.
- [66] R Rogier, H Evans-Marin, J Manasson, et al., Alteration of the intestinal microbiome characterizes preclinical inflammatory arthritis in mice and its modulation attenuates established arthritis. *Sci Rep* 7 (1) (2017) 15613, doi:10.1038/s41598-017-15802-x.
- [67] JU Scher, A Sczesnak, RS Longman, et al., Expansion of intestinal Prevotella copri correlates with enhanced susceptibility to arthritis. *Elife* 2 (2013) e01202, doi:10.7554/eLife.01202.
- [68] X Zhang, D Zhang, H Jia, et al., The oral and gut microbiomes are perturbed in rheumatoid arthritis and partly normalized after treatment. *Nat Med* 21 (8) (2015) 895–905, doi:10.1038/nm.3914.
- [69] Y Maeda, T Kurakawa, E Umemoto, et al., Dysbiosis Contributes to Arthritis Development via Activation of Autoreactive T Cells in the Intestine. *Arthritis Rheumatol* 68 (11) (2016) 2646–2661, doi:10.1002/art.39783.
- [70] J Chen, K Wright, JM Davis, et al., An expansion of rare lineage intestinal microbes characterizes rheumatoid arthritis. *Genome Med* 8 (1) (2016) 43, doi:10.1186/s13073-016-0299-7.
- [71] Y Maeda, T Kurakawa, E Umemoto, et al., Dysbiosis Contributes to Arthritis Development via Activation of Autoreactive T Cells in the Intestine. *Arthritis Rheumatol* 68 (11) (2016) 2646–2661, doi:10.1002/art.39783.
- [72] WK Jubair, JD Hendrickson, EL Severs, et al., Modulation of Inflammatory Arthritis in Mice by Gut Microbiota Through Mucosal Inflammation and Autoantibody Generation. *Arthritis Rheumatol* 70 (8) (2018) 1220–1233, doi:10.1002/art.40490.
- [73] F Sommer, F Bäckhed, The gut microbiota—masters of host development and physiology. *Nat Rev Microbiol* 11 (4) (2013) 227–238, doi:10.1038/nrmicro2974.



- [74] SL Sanos, VL Bui, A Mortha, et al., RORgammat and commensal microflora are required for the differentiation of mucosal interleukin 22-producing Nkp46+ cells, *Nat Immunol* 10 (1) (2009) 83–91, doi:10.1038/ni.1684.
- [75] FM Pavel, CM Vesa, G Gheorghie, et al., Highlighting the Relevance of Gut Microbiota Manipulation in Inflammatory Bowel Disease, *Diagnostics* (Basel) 11 (6) (2021) 1090, doi:10.3390/diagnostics11061090.
- [76] A. Fasano, All disease begins in the (leaky) gut: role of zonulin-mediated gut permeability in the pathogenesis of some chronic inflammatory diseases, *F1000Res*. 9 (2020) F1000 Faculty Rev-69, doi:10.12688/f1000research.20510.1.
- [77] C Sturgeon, A. Fasano, Zonulin, a regulator of epithelial and endothelial barrier functions, and its involvement in chronic inflammatory diseases, *Tissue Barriers* 4 (4) (2016) e1251384, doi:10.1080/21688370.2016.1251384.
- [78] JM Bae, JY Choo, KJ Kim, et al., Association of inflammatory bowel disease with ankylosing spondylitis and rheumatoid arthritis: A nationwide population-based study, *Mod Rheumatol* 27 (3) (2017) 435–440, doi:10.1080/14397595.2016.1211229.
- [79] LX Aa, F Fei, Q Qi, et al., Rebalancing of the gut flora and microbial metabolism is responsible for the anti-arthritis effect of kaempferol, *Acta Pharmacol Sin* 41 (1) (2020) 73–81, doi:10.1038/s41401-019-0279-8.
- [80] J Ren, B Yue, H Wang, et al., Acacetin Ameliorates Experimental Colitis in Mice via Inhibiting Macrophage Inflammatory Response and Regulating the Composition of Gut Microbiota, *Front Physiol* 11 (2021) 577237, doi:10.3389/fphys.2020.577237.
- [81] L Zhu, LZ Xu, S Zhao, et al., Protective effect of baicalin on the regulation of Treg/Th17 balance, gut microbiota and short-chain fatty acids in rats with ulcerative colitis, *Appl Microbiol Biotechnol* 104 (12) (2020) 5449–5460, doi:10.1007/s00253-020-10527-w.
- [82] M Wu, P Li, Y An, et al., Phloretin ameliorates dextran sulfate sodium-induced ulcerative colitis in mice by regulating the gut microbiota, *Pharmacol Res* 150 (2019) 104489, doi:10.1016/j.phrs.2019.104489.
- [83] B Yue, J Ren, Z Yu, et al., Pinocembrin alleviates ulcerative colitis in mice via regulating gut microbiota, suppressing TLR4/MD2/NF- $\kappa$ B pathway and promoting intestinal barrier, *Biosci Rep* 40 (7) (2020) BSR20200986, doi:10.1042/BSR20200986.
- [84] T Behl, K Mehta, A Sehgal, et al., Exploring the role of polyphenols in rheumatoid arthritis, *Crit Rev Food Sci Nutr* (2021) 1–22, doi:10.1080/10408398.2021.1924613.
- [85] D Mauro, F Macaluso, S Fasano, et al., ILC3 in Axial Spondyloarthritis: the Gut Angle, *Curr Rheumatol Rep* 21 (7) (2019) 37, doi:10.1007/s11926-019-0834-9.
- [86] E Chun, S Lavoie, D Fonseca-Pereira, et al., Metabolite-Sensing Receptor Ffar2 Regulates Colonic Group 3 Innate Lymphoid Cells and Gut Immunity, *Immunity* 51 (5) (2019) 871–884.e6, doi:10.1016/j.immuni.2019.09.014.
- [87] A Takaki-Kuwahara, Y Arinobu, K Miyawaki, et al., CCR6+ group 3 innate lymphoid cells accumulate in inflamed joints in rheumatoid arthritis and produce Th17 cytokines, *Arthritis Res Ther* 21 (1) (2019) 198 Published 2019 Aug 30, doi:10.1186/s13075-019-1984-x.
- [88] J Ren, Z Feng, Z Lv, et al., Natural killer-22 cells in the synovial fluid of patients with rheumatoid arthritis are an innate source of interleukin 22 and tumor necrosis factor- $\alpha$ , *J Rheumatol* 38 (10) (2011) 2112–2118, doi:10.3899/jrheum.101377.
- [89] A Toubal, I Nel, S Lotersztajn, et al., Mucosal-associated invariant T cells and disease, *Nat Rev Immunol* 19 (10) (2019) 643–657, doi:10.1038/s41577-019-0191-y.
- [90] M Kim, SJ Yoo, SW Kang, et al., TNF $\alpha$  and IL-1 $\beta$  in the synovial fluid facilitate mucosal-associated invariant T (MAIT) cell migration, *Cytokine* 99 (2017) 91–98, doi:10.1016/j.cyto.2017.07.007.
- [91] Crotty S.T Follicular, Helper Cell Biology: A Decade of Discovery and Diseases, *Immunity* 50 (5) (2019) 1132–1148, doi:10.1016/j.immuni.2019.04.011.
- [92] S Nowotschin, AK Hadjantonakis, Use of KikGR a photoconvertible green-to-red fluorescent protein for cell labeling and lineage analysis in ES cells and mouse embryos, *BMC Dev Biol* 9 (2009) 49, doi:10.1186/1471-213X-9-49.
- [93] G Cao, S Chi, X Wang, et al., CD4+CXCR5+PD-1+ T Follicular Helper Cells Play a Pivotal Role in the Development of Rheumatoid Arthritis, *Med Sci Monit* 25 (2019) 3032–3040, doi:10.12659/MSM.914868.
- [94] J Yang, X Yang, J Yang, et al., Baicalin ameliorates lupus autoimmunity by inhibiting differentiation of Tfh cells and inducing expansion of Tfr cells, *Cell Death Dis* 10 (2) (2019) 140, doi:10.1038/s41419-019-1315-9.
- [95] FB Feng, HY. Qiu, Effects of Artesunate on chondrocyte proliferation, apoptosis and autophagy through the PI3K/AKT/mTOR signaling pathway in rat models with rheumatoid arthritis, *Biomed Pharmacother* 102 (2018) 1209–1220, doi:10.1016/j.biopha.2018.03.142.
- [96] EK Kim, EJ. Choi, Pathological roles of MAPK signaling pathways in human diseases, *Biochim Biophys Acta* 1802 (4) (2010) 396–405, doi:10.1016/j.bbdis.2009.12.009.
- [97] H Du, X Zhang, Y Zeng, et al., A Novel Phytochemical, DIM, Inhibits Proliferation, Migration, Invasion and TNF- $\alpha$  Induced Inflammatory Cytokine Production of Synovial Fibroblasts From Rheumatoid Arthritis Patients by Targeting MAPK and AKT/mTOR Signal Pathway, *Front Immunol* 10 (2019) 1620, doi:10.3389/fimmu.2019.01620.
- [98] G Sun, C Xing, L Zeng, et al., Flemingia philippinensis Flavonoids Relieve Bone Erosion and Inflammatory Mediators in CIA Mice by Downregulating NF- $\kappa$ B and MAPK Pathways, *Mediators Inflamm* 2019 (2019) 5790291, doi:10.1155/2019/5790291.
- [99] G Sun, C Xing, L Zeng, et al., Flemingia philippinensis Flavonoids Relieve Bone Erosion and Inflammatory Mediators in CIA Mice by Downregulating NF- $\kappa$ B and MAPK Pathways, *Mediators Inflamm* 2019 (2019) 5790291, doi:10.1155/2019/5790291.
- [100] KF Zhai, H Duan, CY Cui, et al., Liquiritin from Glycyrrhiza uralensis Attenuating Rheumatoid Arthritis via Reducing Inflammation, Suppressing Angiogenesis, and Inhibiting MAPK Signaling Pathway, *J Agric Food Chem* 67 (10) (2019) 2856–2864, doi:10.1021/acs.jafc.9b00185.
- [101] Y Bao, YW Sun, J Ji, et al., Genkwanin ameliorates adjuvant-induced arthritis in rats through inhibiting JAK/STAT and NF- $\kappa$ B signaling pathways, *Phytomedicine* 63 (2019) 153036, doi:10.1016/j.phymed.2019.153036.
- [102] L Lou, Y Liu, J Zhou, et al., Chlorogenic acid and luteolin synergistically inhibit the proliferation of interleukin-1 $\beta$ -induced fibroblast-like synoviocytes through regulating the activation of NF- $\kappa$ B and JAK/STAT-signaling pathways, *Immunopharmacol Immunotoxicol* 37 (6) (2015) 499–507, doi:10.3109/08923973.2015.1095763.
- [103] L Bai, Y Bai, Y Yang, et al., Baicalin alleviates collagen-induced arthritis and suppresses TLR2/MyD88/NF- $\kappa$ B p65 signaling in rats and HFLS-RAs, *Mol Med Rep* 22 (4) (2020) 2833–2841, doi:10.3892/mmr.2020.11369.
- [104] J Chu, X Wang, H Bi, et al., Dihydropyridinone relieves rheumatoid arthritis symptoms and suppresses expression of pro-inflammatory cytokines via the activation of Nrf2 pathway in rheumatoid arthritis model, *Int Immunopharmacol* 59 (2018) 174–180, doi:10.1016/j.intimp.2018.04.001.
- [105] W Li, TO Khor, C Xu, et al., Activation of Nrf2-antioxidant signaling attenuates NFKappaB-inflammatory response and elicits apoptosis, *Biochem Pharmacol* 76 (11) (2008) 1485–1489, doi:10.1016/j.bcp.2008.07.017.
- [106] TO Khor, MT Huang, KH Kwon, et al., Nrf2-deficient mice have an increased susceptibility to dextran sulfate sodium-induced colitis, *Cancer Res* 66 (24) (2006) 11580–11584, doi:10.1158/0008-5472.CAN-06-3562.
- [107] A Karatas, AF Dagli, C Orhan, et al., Epigallocatechin 3-gallate attenuates arthritis by regulating Nrf2, HO-1, and cytokine levels in an experimental arthritis model, *Biotechnol Appl Biochem* 67 (3) (2020) 317–322, doi:10.1002/bab.1860.
- [108] X Su, Q Huang, J Chen, et al., Calycosin suppresses expression of pro-inflammatory cytokines via the activation of p62/Nrf2-linked heme oxygenase 1 in rheumatoid arthritis synovial fibroblasts, *Pharmacol Res* 113 (Pt A) (2016) 695–704, doi:10.1016/j.phrs.2016.09.031.
- [109] M Zimmermann, M Zimmermann-Kogadeeva, R Wegmann, AL Goodman, Mapping human microbiome drug metabolism by gut bacteria and their genes, *Nature* 570 (7762) (2019) 462–467, doi:10.1038/s41586-019-1291-3.
- [110] L Wang, Q. Ma, Clinical benefits and pharmacology of scutellarin: A comprehensive review, *Pharmacol Ther* 190 (2018) 105–127, doi:10.1016/j.pharmthera.2018.05.006.

## Further reading

- KT Tang, CC Lin, SC Lin, et al., Kurarione Attenuates Collagen-Induced Arthritis in Mice by Inhibiting Th1/Th17 Cell Responses and Oxidative Stress, *Int J Mol Sci* 22 (8) (2021) 4002, doi:10.3390/ijms22084002.
- L Wang, N Wang, Q Zhao, et al., Pectolinarin inhibits proliferation, induces apoptosis, and suppresses inflammation in rheumatoid arthritis fibroblast-like synoviocytes by inactivating the phosphatidylinositol 3 kinase/protein kinase B pathway, *J Cell Biochem* 120 (9) (2019) 15202–15210, doi:10.1002/jcb.28784.